

## Genomic DNA Isolation Kit (Blood/Cultured Cell/Fungus)

Cat No. PDC09-0100(S)

Size: 100 Reactions



**Sample:** Up to 300 µl of the whole blood  
Up to 200 µl of the frozen blood  
Up to 200 µl of the buffy coat  
Cultured animal cells (up to  $1 \times 10^7$ )  
Cultured bacterial cells (up to  $1 \times 10^9$ )  
Fungus cells (up to  $5 \times 10^7$ )

**Format:** Reagent and spin column  
**Yield:** Up to 50 µg  
**Operation time:** Within 60 minutes  
**Elution volume:** 50~100 µl

### Description

The **Genomic DNA Isolation Kit (Blood/Cultured Cell/Fungus)** is designed specifically for genomic DNA isolation from the whole blood, frozen blood, buffy coat, cultured animal/bacterial cells and fungus. This unique buffer system ensures genomic DNA with high yield and good quality from samples. The spin column is designed to purify or concentrate genomic DNA products which have been previously isolated using buffers. The entire procedure can be completed in 1 hour without phenol/ chloroform extraction. Purified genomic DNA is suitable for use in PCR or other enzymatic reactions.

### Feature

- Delivering high-quality genomic DNA with the fast procedure
- Ready-to-use genomic DNA for high performance in any downstream application
- Highly purified and high yield genomic DNA can be extracted from various samples
- Optimized lysis buffer for the efficient lysis
- Designed to rapidly purify high-quality DNA using spin column format

### Kit Contents

Contents	PDC09-0100	PDC09-0100S
Buffer CR	100 ml	4 ml
Buffer CC	35 ml	1.5 ml
Buffer CB	45 ml	2 ml
Buffer W1	45 ml	2 ml
Buffer W2 (Add ethanol)	15 ml (60 ml)	300 µl x2 (1.2 ml x2)
Buffer RE	10 ml	1 ml
Columns CC	100 pcs	4 pcs
Collection Tubes	100 pcs	4 pcs

### Quality Control

The quality of the Genomic DNA Isolation Kit (Blood/Cultured Cell/Fungus) is tested on a lot-to-lot basis to ensure consistent product quality.

### Required Materials

- 1.5 ml Microcentrifuge tubes
- lysozyme buffer (for Gram-positive Bacteria):  
20 mg/ml lysozyme; 20 mM Tris-HCl; 2 mM EDTA; 1% Triton X-100; pH 8.0
- Lyticase or zymolase (for fungus)
- sorbitol buffer (for fungus):  
1.2 M sorbitol; 10 mM CaCl<sub>2</sub>; M Tris-HCl pH 7.5; 35 mM β- mercaptoethanol
- RNase A (10 mg/ml)
- Absolute ethanol (96~100%)

## Genomic DNA Isolation Kit Protocol Fresh whole Blood or Buffy Coat

### Step 1 Sample Cells Harvesting

1. Collect blood in EDTA-Naz treated collection tubes (or other anticoagulant mixtures).
2. Transfer up to 300 µl of the blood or 200 µl of the buffy coat to a sterile 1.5 ml microcentrifuge tube.
3. Add 900 µl of the Buffer CR and mix by inversion.
4. Incubate the tube at the room temperature for 10 minutes (invert twice during incubation).
5. Centrifuge for 5 minutes at 4,000 x g.
6. Remove the supernatant completely and resuspend the cells in 50 µl of the Buffer CR by pipetting the pellet.

### Step 2 Lysis

1. Add 300 µl of the Buffer CC to the resuspended cells from Step 1 and mix by vortex.
2. Incubate at 60°C for 10 minutes or until the sample lysate is clear. During incubation, invert the tube every 3 minutes.

### Optional Step:

- ◆ RNA Degradation (If RNA-free genomic DNA is required, perform this optional step)
3. Add 5 µl of the RNase A (10 mg/ml) to the sample lysate and mix by vortex. Incubate at the room temperature for 5 minutes.

### Step 3 Protein Removal

1. Add 400 µl of the Buffer CB to the sample from Step 2 and shake vigorously.
2. Centrifuge at 12,000 x g for 1 minute. (Don't over 1 minute)

### Step 4 DNA Binding

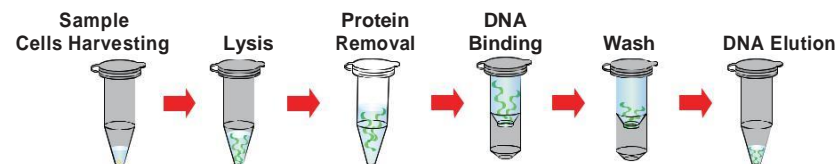
1. Place a Column CC in a 2 ml Collection Tube.
2. Transfer the clear supernatant completely from the previous step to the Column CC.
3. Centrifuge at 14,000 x g for 30 seconds.
4. Discard the flow-through and place the Column CC back in the same Collection Tube.

### Step 5 Wash

1. Add 400 µl of the Buffer W1 into the Column CC.
2. Centrifuge at 14,000 x g for 30 seconds.
3. Discard the flow-through and place the Column CC back in the same Collection Tube.
4. Add 600 µl of the Buffer W2 (Ethanol added) into the Column CC.
5. Centrifuge at 14,000 x g for 30 seconds.
6. Discard the flow-through and place the Column CC back in the same Collection Tube.
7. Centrifuge at 14,000 x g again for 2 minutes to remove residual Buffer W2.

### Step 6 DNA Elution

1. Transfer the dried Column CC to a new 1.5 ml microcentrifuge tube.
2. Add 50-100 µl of the Pre-Heated Buffer RE or TE into the center of the column matrix.
3. Let stand at 60°C for 3 minutes.
4. Centrifuge for 2 minutes at 14,000 x g to elute the purified DNA.



## Genomic DNA Isolation Kit Protocol Cultured Mammalian Cells

### Step 1 Sample Cells Harvesting

1. Transfer cultured mammalian cells (up to  $10^7$ ) to a sterile 1.5 ml microcentrifuge tube.
2. Centrifuge at 6,000 x g for 1 minute.
3. Remove the supernatant completely and resuspend the cells in 50  $\mu$ l of the Buffer CR by pipetting the pellet.

### Step 2 Lysis

1. Add 300  $\mu$ l of the Buffer CC to the resuspended cells from Step 1 and mix by vortex.
2. Incubate at 60°C for 10 minutes or until the sample lysate is clear. During incubation, invert the tube every 3 minutes.

#### Optional Step:

- ◆ RNA Degradation (If RNA-free genomic DNA is required, perform this optional step)
3. Add 5  $\mu$ l of the RNase A (10 mg/ml) to the sample lysate and mix by vortex. Incubate at the room temperature for 5 minutes.

### Step 3 Protein Removal

1. Add 400  $\mu$ l of the Buffer CB to the sample from Step 2 and shake vigorously.
2. Centrifuge at 12,000 x g for 1 minute. (Don't over 1 minute)

### Step 4 DNA Binding

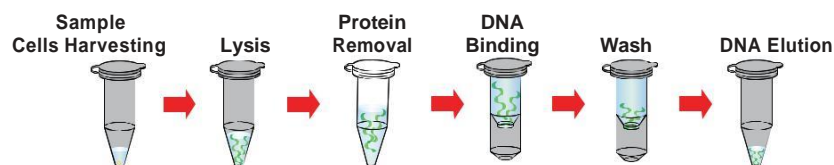
1. Place a Column CC in a 2 ml Collection Tube.
2. Transfer the clear supernatant completely from the previous step to the Column CC.
3. Centrifuge at 14,000 x g for 30 seconds.
4. Discard the flow-through and place the Column CC back in the same Collection Tube.

### Step 5 Wash

1. Add 400  $\mu$ l of the Buffer W1 into the Column CC.
2. Centrifuge at 14,000 x g for 30 seconds.
3. Discard the flow-through and place the Column CC back into the same Collection tube.
4. Add 600  $\mu$ l of the Buffer W2 (Ethanol added) into the Column CC.
5. Centrifuge at 14,000 x g for 30 seconds.
6. Discard the flow-through and place the Column CC back into the same Collection tube.
7. Centrifuge at 14,000 x g again for 2 minutes to remove residual Buffer W2.

### Step 6 DNA Elution

1. Transfer the dried Column CC to a new 1.5 ml microcentrifuge tube.
2. Add 50-100  $\mu$ l of the Pre-Heated Buffer RE or TE into the center of the column matrix.
3. Let stand at 60°C for 3 minutes.
4. Centrifuge for 2 minutes at 14,000 x g to elute the purified DNA.



## Genomic DNA Isolation Kit Protocol Gram-Negative Bacterial Cells

### Step 1 Sample Cells Harvesting

1. Transfer cultured bacterial cells (up to  $10^9$ ) to a sterile 1.5 ml microcentrifuge tube.
2. Centrifuge at 12,000 x g for 1 minute.
3. Remove the supernatant completely and resuspend the cells in 50  $\mu$ l of Buffer CR by pipetting the pellet.

### Step 2 Lysis

1. Add 300  $\mu$ l of Buffer CC to the resuspended cells from Step 1 and mix by vortex.
2. Incubate at 60°C for 10 minutes or until the sample lysate is clear. During incubation, invert the tube every 3 minutes.

#### Optional Step:

- ◆ RNA Degradation (If RNA-free genomic DNA is required, perform this optional step)
3. Add 5  $\mu$ l of the RNase A (10 mg/ml) to the sample lysate and mix by vortex. Incubate at the room temperature for 5 minutes.

### Step 3 Protein Removal

1. Add 400  $\mu$ l of the Buffer CB to the sample from Step 2 and shake vigorously.
2. Centrifuge at 12,000 x g for 1 minute. (Don't over 1 minute)

### Step 4 DNA Binding

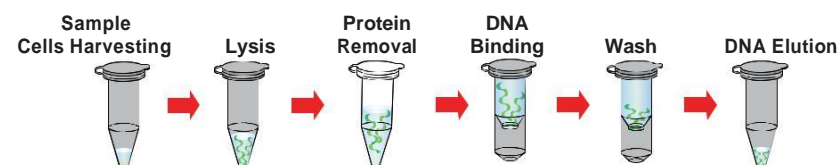
1. Place a Column CC in a 2 ml Collection Tube.
2. Transfer the clear supernatant completely from the previous step to the Column CC.
3. Centrifuge at 14,000 x g for 30 seconds.
4. Discard the flow-through and place the Column CC back in the same Collection Tube.

### Step 5 Wash

1. Add 400  $\mu$ l of the Buffer W1 into the Column CC.
2. Centrifuge at 14,000 x g for 30 seconds.
3. Discard the flow-through and place the Column CC back into the same Collection tube.
4. Add 600  $\mu$ l of the Buffer W2 (Ethanol added) into the Column CC.
5. Centrifuge at 14,000 x g for 30 seconds.
6. Discard the flow-through and place the Column CC back into the same Collection tube.
7. Centrifuge at 14,000 x g again for 2 minutes to remove the residual Buffer W2.

### Step 6 DNA Elution

1. Transfer the dried Column CC to a new 1.5 ml microcentrifuge tube.
2. Add 50-100  $\mu$ l of Pre-Heated Buffer RE or TE into the center of the column matrix.
3. Let stand at 60°C for 3 minutes.
4. Centrifuge for 2 minutes at 14,000 x g to elute the purified DNA.



## Genomic DNA Isolation Kit Protocol Gram-Positive Bacterial Cells

### Step 1 Sample Cells Harvesting

1. Transfer cultured bacterial cells (up to  $10^9$ ) to a sterile 1.5 ml microcentrifuge tube.
2. Centrifuge at 12,000 x g for 1 minute.
3. Remove the supernatant completely and resuspend the cells in 100  $\mu$ l of lysozyme buffer by pipetting the pellet. Incubate at room temperature for 20 minutes.

### Step 2 Lysis

1. Add 300  $\mu$ l of Buffer CC to the resuspended cells from Step 1 and mix by vortex.
2. Incubate at 60°C for 10 minutes or until the sample lysate is clear. During incubation, invert the tube every 3 minutes.

#### Optional Step:

- ◆ RNA Degradation (If RNA-free genomic DNA is required, perform this optional step)
- 3. Add 5  $\mu$ l of the RNase A (10 mg/ml) to the sample lysate and mix by vortex. Incubate at the room temperature for 5 minutes.

### Step 3 Protein Removal

1. Add 400  $\mu$ l of Buffer CB to the sample from Step 2 and shake vigorously.
2. Centrifuge at 12,000 x g for 1 minute. (Don't over 1 minute)

### Step 4 DNA Binding

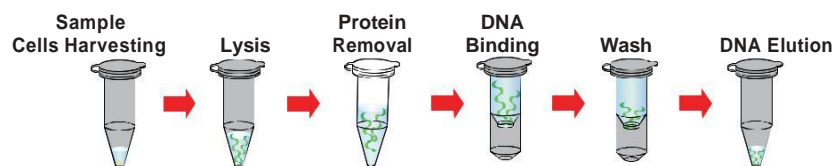
1. Place a Column CC in a 2 ml Collection Tube.
2. Transfer the clear supernatant completely from the previous step to the Column CC.
3. Centrifuge at 14,000 x g for 30 seconds.
4. Discard the flow-through and place the Column CC back in the same Collection Tube.

### Step 5 Wash

1. Add 400  $\mu$ l of Buffer W1 into the Column CC.
2. Centrifuge at 14,000 x g for 30 seconds.
3. Discard the flow-through and place the Column CC back into the same Collection tube.
4. Add 600  $\mu$ l of Buffer W2 (Ethanol added) into the Column CC.
5. Centrifuge at 14,000 x g for 30 seconds.
6. Discard the flow-through and place the Column CC back into the same Collection tube.
7. Centrifuge at 14,000 x g again for 2 minutes to remove the residual Buffer W2.

### Step 6 DNA Elution

1. Transfer the dried Column CC to a new 1.5 ml microcentrifuge tube.
2. Add 50-100  $\mu$ l of Pre-Heated Buffer RE or TE into the center of the column matrix.
3. Let stand at 60°C for 3 minutes.
4. Centrifuge for 2 minutes at 14,000 x g to elute the purified DNA.



## Genomic DNA Isolation Kit Protocol Fungus Cells

### Step 1 Sample Cells Harvesting

1. Transfer fungus cells (up to  $10^8$ ) to a sterile 1.5 ml microcentrifuge tube.
2. Centrifuge at 6,000 x g for 5 minute.
3. Remove the supernatant completely and resuspend the cells in 600  $\mu$ l of sorbitol buffer by pipetting the pellet.
4. Add 200 U of the lyticase or zymolase. Incubate at 30°C for 30 minutes.
5. Centrifuge the mixture for 10 minutes at 2,000 x g to harvest the spheroplast. Remove the supernatant completely and resuspend the cells in 50  $\mu$ l of the Buffer CR by pipetting the pellet.

### Step 2 Lysis

1. Add 300  $\mu$ l of the Buffer CC to the resuspended cells from Step 1 and mix by vortex.
2. Incubate at 60°C for 10 minutes or until the sample lysate is clear. During incubation, invert the tube every 3 minutes.

#### Optional Step:

- ◆ RNA Degradation (If RNA-free genomic DNA is required, perform this optional step)
- 3. Add 5  $\mu$ l of the RNase A (10 mg/ml) to the sample lysate and mix by vortex. Incubate at the room temperature for 5 minutes.

### Step 3 Protein Removal

1. Add 400  $\mu$ l of the Buffer CB to the sample from Step 2 and shake vigorously.
2. Centrifuge at 12,000 x g for 1 minute. (Don't over 1 minute)

### Step 4 DNA Binding

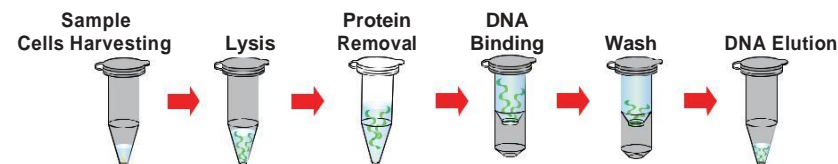
1. Place a Column CC in a 2 ml Collection Tube.
2. Transfer the clear supernatant completely from the previous step to the Column CC.
3. Centrifuge at 14,000 x g for 30 seconds.
4. Discard the flow-through and place the Column CC back in the same Collection Tube.

### Step 5 Wash

1. Add 400  $\mu$ l of the Buffer W1 into the Column CC.
2. Centrifuge at 14,000 x g for 30 seconds.
3. Discard the flow-through and place the Column CC back into the same Collection tube.
4. Add 600  $\mu$ l of the Buffer W2 (Ethanol added) into the Column CC.
5. Centrifuge at 14,000 x g for 30 seconds.
6. Discard the flow-through and place the Column CC back into the same Collection tube.
7. Centrifuge at 14,000 x g again for 2 minutes to remove the residual Buffer W2.

### Step 6 DNA Elution

1. Transfer the dried Column CC to a new 1.5 ml microcentrifuge tube.
2. Add 50-100  $\mu$ l of the Pre-Heated Buffer RE or TE into the center of the column matrix.
3. Let stand at 60°C for 3 minutes.
4. Centrifuge for 2 minutes at 14,000 x g to elute the purified DNA.



## Troubleshooting

Refer to the table below to troubleshoot problems that you may encounter when purifying the genomic DNA with the kit.

Problem	Cause	Solution
Low yield of DNA	Incomplete lysed sample	Use the appropriate method for the lysate preparation based on the amount of the starting materials.
		Increase the digestion time.
DNA degrade	Sample not fresh	Avoid repeated freeze / thaw cycles of the sample.
		Use a new sample for the DNA isolation. Perform the extraction of the fresh material when possible.
	Inappropriate sample storage conditions	Store culture cell / fungus at -20°C until use. The whole blood can be stored at 4°C for no longer than 1~2 days.
	DNase contaminant	Use the fresh TAE or TBE electrophoresis buffer.
		Maintain a sterile work environment to avoid contamination from DNases.
Presence of RNA	RNA contamination	Perform RNase A digestion step during Step Lysis.
Inhibition of downstream enzymatic reactions	Presence of ethanol in purified DNA	Discard the ethanol of the Buffer W2 flow through from the collection tube. Place the spin cartridge into the collection tube and centrifuge the spin cartridge at maximum speed for 2– 3 minutes to completely dry the cartridge.

## Caution:

- During operation, always wear a lab coat, disposable gloves, and protective equipment.
- Check buffers before use for salt precipitation. Redissolve any precipitate by warming to 37°C.
- Add 60 ml of the ethanol (96–100%) to the Buffer W2, and shake before use.
- Buffers W1 contain irritants. Wear gloves when handling these buffers.
- Research Use Only. Not intended for any animal or human therapeutic or diagnostic uses.