# PanGreen™ Universal SYBR® Green Master Mix

28 JUL 2025

| Catalog Number | Size | Concentration |
|---|---|---|
| QSD01-0100 | 100 reactions (20 µl vol) | 2X |

## Storage Conditions

Stable for up to 3 months at 4°C.

Stable for up to 24 months at -20°C.

## Description

PanGreen™ Universal SYBR® Green Master Mix is a 2x concentrated, ready for use Master Mix reaction enhanced for dye-based quantitative PCR (qPCR) and compatible with the majority of commercially available real-time PCR systems (ROX-independent and ROX-dependent). It contains NanoTaq hot-start DNA polymerase, dNTPs, $MgCl_2$, SYBR® Green I dye, enhancers, stabilizers and essentials for a success PCR reaction.

## Kit Content(s)

| | |
|---|---|
| 2X universal SYBR® Green Master Mix | 1 ml x 1 vial |
| High ROX Reference Dye | 40 µl x 1 vial |
| Low ROX Reference Dye | 40 µl x 1 vial |

## Required materials but not provided

- A compatible real-time PCR instrument
- Vortex or equivalent
- Microcentrifuge
- Plates and seals for your instruments

## Instrument Compatibility

This Master Mix is compatible with the majority of commercially available real-time PCR systems.

| Instrument | ROX |
|---|---|
| ABI Prism7000/7300/7700/7900HT, ABI Step One, ABI Step One Plus | High ROX reference dye |
| ABI Prism 7500/7500 Fast, MJ Research Chromo4, Option (II), Corbett Rotor Gene 3000 | Low ROX reference dye |

## Reaction Setup

1. Thaw PanGreen™ Universal SYBR® Green Master Mix and the rest of frozen reaction components to a temperature of 4°C. In order to entirely collect solutions, combine thoroughly and centrifuge briefly, then store at 4°C and avoid from light.



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2. Prepare (on ice or at room temperature) enough assay Master Mix for all reactions by adding all necessary components, except the DNA template, according to the recommendations in Table 1 (below).

**Table 1. Reaction Setup**

| Components | Volume per 20 µl Reaction | Volume per 10 µl Reaction | Final Concentration |
|---|---|---|---|
| PanGreen™ Universal SYBR® Green Master Mix (2x) | 10 µl | 5 µl | 1x |
| Forward and reverse primers | Variable | Variable | 300–500 nM each primer |
| DNA template **(add at step 4)** | Variable | Variable | cDNA: 1pg–10ng<br>Genomic DNA: 50ng-250ng |
| High ROX Reference Dye | 0.4 µl | 0.2 µl | 500 nM or not required |
| Low ROX Reference Dye | 0.4 µl | 0.2 µl | 50 nM or not required |
| Nuclease-free $H_2O$ | Variable | Variable | -— |
| *Total reaction mix volume* | *20 µl* | *10 µl* | -— |

* Optimization may be needed for better performance.

3. Combine the assay Master Mix thoroughly to ensure consistency and equally dispense the solution into each qPCR tube or into the wells of a qPCR plate. Employ good pipetting practice to ensure assay precision and accuracy.
4. Add DNA samples (and DNase-free $H_2O$ if needed) to the PCR tubes or wells containing assay Master Mix (Table 1), seal the tubes or wells with flat caps or optically transparent film. **Note:** to ensure thorough mixing of reaction components, vortex for approximately 30 seconds (or more).
5. Spin the tubes or plate to remove any air bubbles and collect the reaction mixture in the vessel bottom.
6. Setup the thermal cycling protocol on a real-time PCR instrument according to Table 2. **Note:** optimization may be needed for better performance.
7. Load the PCR tubes or plate into the real-time PCR instrument and commence the run.
8. Perform data analysis according to the instrument-specific instructions.

- Process in the thermal cycler for 35~45 cycles as follows:

**Table 2. Thermal Cycling Protocol**

| | |
|---|---|
| Initial Denaturation | 3-5 minutes at 95°C (5 mins for GC rich or complex templates) |
| Denaturation | 15 seconds at 95°C |
| Annealing & Extension | 60 seconds at 60°C and Plate Read |
| Melting curve | Refer to specific guidelines for instrument used |

Note: Optimal conditions for amplification will vary depending on the primers and thermal cycler used. It may be necessary to optimize the system for individual primers, template, and thermal cycler.

## Template

Purified high quality DNA is needed for a success PCR reaction. The final concentration of cDNA template please refer to table 1.

## Important notes

1. Shake gently before use to avoid foaming and low-speed centrifugation.
2. During operation, always wear a lab coat, disposable gloves, and protective equipment.

## Troubleshooting

Refer to the table 3 below to troubleshoot problems that you may encounter when quantifying of nucleic acid targets with the kit.

**Table 3. Troubleshooting**

| Trouble | Cause | Solution |
|---|---|---|
| Poor Signal or No Signal | Inhibitor Present | 1. Perform a dilution series of the PCR template to determine whether the effect of the inhibitory agent can be reduced.<br>2. Take extra care with the nucleic acid extraction steps to minimize carryover of PCR inhibitors. |
| | Degraded Template Material | 1. Do not store diluted template in water or at low concentrations.<br>2. Check the integrity of template material by automated or manual gel electrophoresis. |
| | Inadequate Thermal Cycling Conditions | 1. Try using a minimum extension time of 30 sec for genomic DNA and 15 sec for cDNA. |
| Signal in Negative Control | Contamination of Reaction Components with Target Sequence | 1. To minimize the possibility of contamination of PCR components by PCR product or other template, designate a work area exclusively for PCR assay setup.<br>2. Use a solution of 10% bleach instead of ethanol to prepare the workstation area for PCR assay setup. Ethanol will only induce precipitation of DNA in your work area, while the 10% bleach solution will hydrolyze, as well as dissolve, any residual DNA. |
| Poor Reproducibility Across Replicate Samples | Inhibitor Present | 1. Perform a dilution series of the PCR template to determine whether the effect of the inhibitory agent can be reduced.<br>2. Take extra care with the nucleic acid extraction steps to minimize carryover of PCR inhibitors. |
| | Primer Design | 1. Verify primers design at different annealing temperatures. |


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| | | |
|---|---|---|
| Low or High Reaction Efficiency | Primer- Dimer | 1. Reduce primer concentration.<br>2. Evaluate primer sequences for complementarity and secondary structure. Redesign primers if necessary.<br>2. Perform melt-curve analysis to determine if primer- dimers are present. |
| | Insufficient Optimization | 3. Use a thermal gradient to identify the optimal thermal cycling conditions for a specific primer set. |


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